#### THE PROTECTIVE AND CURRATIVE EFFECT OF *FICUS VOGELII* AGAINST TOXICITY INDUCED BY LEAD-ACETATE ON SPLEEN IN ADULT WISTAR RATS

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ABSTRACT: Objective: This study aimed at investigating the protective and currative effect of Ficus vogelii against toxicity induced by lead acetate on spleen in adult Wistar rats. Methodology: Thirty (30) Wistar rats were used for this research and randomly assigned into 6 groups of five rats. Group A (Control group) received normal saline and water ad libitum. B was given lead acetate in a dose of 2mg/kg body weight for 28 days. The currative groups C (low dose) and D (high dose) received lead acetate and aqueous extract. The protective groups E (low dose) and F (high dose) received aqueous extract and lead acetate. Group G were given only aqueous extract. The experiment lasted for 28 days after 7 days of acclimatization. Results: Figure 1C showed lymphoid follicle with little necrosis demonstrating the effects of the extract as currative herb. Figure 1D shows expansion of red pulp and macrophages with relative improvement when compared to positive control. Figure 3E shows lymphoid follicle with necrosis and affected trabeculae (T). Figure 3F showed diffused red pulp and necrotized macrophages (arrows) which are not as much as that found in the positive control group and may be attributed to the protective role of the extract. The extract only (Figure 4) group shows normal histoarchitecture with red pulp, white pulp, microphages, trabeculae and lymphoid follicle all intact without any injury. Conclusion: This experiment demonstrates that lead causes splenic alterations like diffusion of white pulp into red pulp and appearance of large macrophages due to inflammations and production of debris of dead cells. These alterations can either be ameliorated or protect against by Ficus vogelii extract.

KEYWORDS: Currative, Ficus Vogelii, Lead-Acetate, Protective, Toxicity, Spleen.

## INTRODUCTION

Lead is an important metal with a bluish green colour. It is naturally available in small quantity as part of the earth's crust [1]. Lead is present in almost all parts of our environment such as water, air, food, soil and even fruits. Lead has found in the home for a very long time. Currently, it is used in the manufacturing of light-tech products for protections of nuclear reactors, thin sheets of electronic components, batteries, paints, ceramics, cables and ammunitions [2]. These products in our homes do not only cause health hazards to the people using them alone but due to their emissions, they pollute the environment causing health hazard to other inhabitants.

Lead gains entrance through various routes into the body via nasal, oral or dermal routes and cause intoxication [3,4]. It is very deleterious to almost all organs in the body and has

significant debilitating effects on the nervous, renal, hepatic, circulatory and hematopoietic system [5,6] while the two commonest routes of entry of lead into the body are inhalation and ingestion [7]. Ingestion is seeming to be more common source and inhalation is the most significant because pulmonary absorption is more efficient. They are various organs in the body that also show critical damage [8,9]. The ingested and absorbed lead are stored primarily in soft tissues and bone but the highest concentration of lead occurs in the bones, teeth, liver, lung, kidney, brain and spleen [10].

The spleen is the largest lymphoid organ with a very rich blood supply. The spleen is surrounded by a connective tissue capsule and divides its interior into incomplete compartments called the splenic pulp [11]. Being an important organ in the body in various ways, the spleen is concerned with phagocytosis and immune responses. In the fetus it is also an important site of haemopoiesis. Postnatally, it may become hematopoietic in some diseased conditions. Although very important in the defense of the body, it may not absolutely be essential because if the spleen is removed, many of its functions can be assumed by the liver and other lymphoid tissues [12,13]. With lead being so invasive, anything damage caused by the lead to the spleen is bound to affect all other organs that are supposed to take up the functions [14,15].

A lot of works has looked at the deleterious/toxic effects of lead on the histoarchitecture of spleen [16,17] and effects of other herbs and other substances on reducing the toxicity [18,19] but no researcher has ever considered the effect of *Ficus vogelii*. It is in view of the above that we designed this research work to evaluate the protective and currative effects of *F*.vogelii on the toxicity induced by lead acetate in the spleen of adult albino Wistar rats.

## MATERIALS AND METHODS

## **Experimental Protocol**

The thirty (30) healthy adult Wistar rats used for this research weighed between 180-200gm. They were acclimatized for 7 days and randomly assigned into 6 groups of five rats each. Group A (Control group) received normal saline and water *ad libitum*. Group B (experimental group) were given lead acetate in a dose of 2mg/kg body weight for 28 days. Group C (currative low dose) were given lead acetate and 100mg/kg of aqueous extract for 28 days. Group D (currative high dose) were given lead acetate and 300mg/kg of aqueous extract for 28 days. Group E (protective low dose) were given 100mg/kg of aqueous extract and lead acetate for 28 days. Group F (protective high dose) were given 300mg/k of aqueous extract only for the same period. All the animals involved in this research received oral administration using orogastric tube [20].

### **Ethical Clearance**

During this experiment, we respected and strictly observed the following councils' directive on the use of experimental animals. They are; Directive 2010/63/Eu, 2010 of the European parliament and the European Council as passed on 22 September, 2010 on the use and protection of experimental animals [21] and the Organization of Economic Co-operation Development, Paris, guideline for testing of chemical usage in Experimental animals, OECD,

[22]. These animals were fed with standard rat feed and allowed water *ad libitum* [19,23]. We also sort for and obtained the ethical clearance from the Faculty of Basic Medical Sciences University of Nigeria Enugu Campus.

### **Leaves Collection and Extraction**

The fresh leaves of *F. vogelii* were collected from Enyibichiri Ndufu-Alike Ikwo in Ikwo Local Government Area of Ebonyi State. These leaves were washed and dried in a ventilated room. Thereafter, it was crushed into powder using electronic blender and passed through mesh sieve to get the fine powders. Five hundred grams (500g) of the powder was weighed using an electronic weighing balance and soaked in 1200mL of water (powder/solvent). The mixture was agitated using an electric blender to enhance proper mixing of the solvent with the powder and then poured into air-tight plastic containers. The container with the mixture was kept in a refrigerator for 48hours [24]. The mixtures were filtered first with cheese cloth, and then with Whiteman No 1 filter paper (24cm). The filtrates were separated and concentrated in vacuum using Rotary Evaporator to 10% of their original volumes at  $37^{0}$ C - 40°C. These were concentrated using a water bath until a sticky paste was gotten. The extracts were stored in a refrigerator until it is required for use. All preparations were performed at the Department of Anatomy Faculty of Basic Medical Sciences, College of Medicine University of Nigeria Enugu Campus (UNEC).

### **Histological Study**

At the end of the experiment, the rats were starved for 24 hours and anaesthetized and then decapitated [25]. The animals were dissected and the liver removed and quickly fixed in 10 % formalin for routine histological procedures. The tissues were processed and embedded in paraffin wax. Thin sections of 4-5 $\mu$ m were obtained and stained using haematoxylin and eosin (H&E) and were examined under light microscope to determine the histological changes in the liver histology.

# RESULTS

The results of the experiment are presented below as a photomicrographic plate. These sections were taken after a very routine histological process had been undertaken.



Figure.1: Photomicrograph of rat; (A) spleen that received only normal saline showing normal white pulp, red pulp and normal spleen histology, H & E, Mag; 200. (B)
Photomicrograph of rat that received only lead acetate showing N-necrotized area, diffused white pulp, distorted red pulp and damaged histology, H & E, Mag; 200.



Figure 2: Photomicrograph of rat; (C) spleen treated with lead acetate before *F.vogelii* 100mg/kg showed lymphoid follicle without necrosis (H&E-Mag, 200). (D) Spleen treated with lead acetate before *F.vogelii* 300mg/kg showed expansion of red pulp and macrophages (arrows) (H&E-Mag, 200).



Figure 3: Photomicrograph of rat; (E) spleen treated with F.vogelii 100mg/kg before showed lymphoid follicle with necrosis and affected trabeculae (T) (H&E-Mag, 200). (F) Spleen treated with F.vogelii 300mg/kg before lead acetate showed diffused of red pulp and necrotized macrophages (arrows) (H&E-Mag, 200).



Figure 4: Photomicrograph of rat spleen that received only 300mg/kg of the aqueous extract showing normal histological architecture with red pulp, white pulp, microphages and lymphoid follicle.

## DISCUSSION

Human health has been in danger recently due to the emissions from our various home appliances as a result of science and technology and from our natural environment. These emissions we may not be in control of but we can either manage the way we interact with them or use our natural products such as the herbs that have been proven to be antitoxicity to counteract their effect. After the present research, the histological investigations of the spleens in the normal control group were not affected as it did not receive anything other than water and feed. All the treated rats showed distortions in their splenic architectonics such as the red and white pulp besides fibrous capsule as a covering for the spleen [26]. According to Aldahmash and El-Nager, [27] the red pulp is the area of spleen in between white pulp and consists of open sinuses and cellular cords. Splenic sinuses are open vascular spaces lined by a discontinuous layer of endothelial cells and supported by a fenestrated basal lamina and reticular fibers. The surrounding cellular splenic cords provide a tissue frame work maintaining the network of sinuses.

The positive control that was treated with lead acetate only showed that the spleen of the rats presented several distortions that represent the deleterious effects of lead on the spleen architecture [28]. The white pulp was caused to diffuse into the red pulp which lead to the two structures being ill defined. There also various areas that showed necrosis as a result of the induced lead acetate toxicity [27]. The cells mostly responsible for slowing the propagation of invading pathogen in the spleen are macrophages and polymorphonuclear (phagocytes cells), these cells are in charge while an antigen-specific adaptive immune response (antibody- or cell-mediated) is being established. Various authors such as Kowolenko *et al.*, [29]; Mauel *et al.*, [30] reported that lead can inhibit macrophage function possibly by overloading macrophages with cellular debris and inhibiting macrophage production of nitric oxide [31].

The photomicrograph of spleen in figure 1C spleen treated with lead acetate before *F.vogelii* 100mg/kg showed lymphoid follicle without much necrosis which may be demonstrating the effects of the extract as currative herb. Figure 1D is showing the spleen of rats treated with lead acetate before *F.vogelii* 300mg/kg with expansion of red pulp and macrophages (arrows) which showed relative improvement when compared with the positive control. Figure 3E represents the spleen of rats treated with *F.vogelii* 100mg/kg before lead acetate showing lymphoid follicle with necrosis and affected trabeculae (T). in figure 3F, the Spleen of the rats treated with *F.vogelii* 300mg/kg before lead acetate showed diffused red pulp and necrotized macrophages (arrows) which are not as much as that found in the positive control group and may be attributed to the protective role of the extract. The results of the present study agreed with those of Aly *et al.* [32] who showed that the effects of lead toxicity in spleen can be countered. Photomicrograph of rat spleen that received only 300mg/kg of the aqueous extract (Figure 4) shows normal histoarchitecture with red pulp, white pulp, microphages, trabeculae and lymphoid follicle all intact without any injury.

Our present experiment demonstrates that oral administration of lead results in several alterations in the spleen represented by diffusion of white pulp into the red pulp and appearance of large macrophages due to the inflammations caused by toxicity and production of debris of dead cells. This agrees with the report of Aldahmash And El-Nager, [27].

### CONCLUSION

The herb did a great work in the restoration of the integrity of the spleen histology that is to say it act well as a currative medicine. The extract was also good as a protective herb. Comparing the both the herb could better be used as a protective medicine because it tends to protect more than when used as currative medicine.

### REFERENCES

- [1] Aldahmash AB and El-Nager DM. Antioxidant effects of captopril against lead acetateinduced hepatic and splenic tissue toxicity in Swiss albino mice. Saudi Journal of Biological Sciences, 2016; 23, 667–673.
- [2] Fioresi M, Simo<sup>e</sup>s MR, Furieri LB, Broseghini-Filho GB, Vescovi MVA, et al. (2014) Chronic Lead Exposure Increases Blood Pressure and Myocardial Contractility in Rats. PLoS ONE 9(5): e96900. doi:10.1371/journal.pone.0096900.
- [3] Türkay M, Türker H, Güven T. Ultrastructural effects of lead acetate on the spleen of rats. *Turk J Biol* (2015) 39:511-516. doi:10.3906/biy-1404-48. Available at ttp://journals.tubitak.gov.tr/biology.
- [4] Nordberg GF, Fowler BA, Nordberg M, Friberg L. Handbook on the Toxicology of Metals. 3rd ed. Amsterdam, Netherlands: Academic Press, 2007; Pg. 1024.
- [5] Siposet P, Szentmihalyi K, Feher E, Abaza M, Szilyagi M and Blazovics A. Some effects of lead ontamination on liver and gall bladder bile. *Acta Biol Szeged* 2003; 47(1-4):139-42.
- [6] Patrick L. Lead toxicity, a review of the literature. Part 1: Exposure, evaluation, and treatment. Alternative medicine review: a journal of clinical therapeutic 2006; 11 (1): 2–22.
- [7] IARC. Summaries & evaluations: Inorganic and organic lead compounds. Lyon, International Agency for Research on Cancer. IARC Monographs for the Evaluation of Carcinogenic Risks to Humans 2006; Vol. 87.
- [8] Khan MH, Mosafa M, Jahan, MS, Sayed MA, Hossain MA. Effects of garlic and vitamin B-complex in lead acetate induced toxicities in mice. Bangl. J. Vet. Med. 6(2), 2008, 203-210.
- [9] Pizzol M, Thomsen M, Anderson M. Long-term human exposure to lead from different media and intake pathways. *Sci. Total Environ.*,2010, 408: 5478-5488.
- [10] Mudipalli A. Lead hepatotoxicity and potential health effects. *Indian J. Med. Res.* 2007; 126, 518–527.
- [11] Eroschenko VO. diFiore's Atlas of Histology with Functional Correlations. 11<sup>th</sup> edition. 2008. Chapt. 9: pg. 191-200. Wolters Kluwer Williams & Wilkins. Philadelphia United Kingdom, London.
- [12] Drake R, Vogl W, Mitchell AWM. Grays Anatomy for Students. 3rd eds, 2014, Chpt, 4: pg. 272-293. Churchill Livingstone (Elsevier's) Publishers; Saunders.
- [13] Standring S, Ellis H, Healy JC, Johnson D, Williams A and Collins P. Gray's Anatomy: The Anatomical Basis of Clinical Practice. 40<sup>th</sup> edition: 2008, Chpt 71, pp1530-1720. Madrid Churchill Livingstone
- [14] Lavicoli I, Carelli G, Stanek EJ. Effects of low doses of dietary lead on red blood cell production in male and female mice. *Toxicol. Lett.* 137, 2003, 193-199.

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- [15] Amirshahrokhi K, Ghazi-khansari M, Farani A, Karimian, G. Effect of captopril on TNF-a and IL-10 in the livers of bile duct ligated rats. *Iran. J. Immunol*, 2010; 7: 247– 251.
- [16] Suradkar S, Vihol P, Patel J, Ghodasara D, Joshi B, Prajapati K. Patho-morphological changes in tissues of Wistar rats by exposure of lead acetate. *Vet. World.* 2010 (3): 82– 84.
- [17] Kamruzzaman. Effects of ascorbic acid (vitamin C) and α-tocophenol (vitamin E) in lead induced toxicities in rats. MS Thesis, Department of Pharmacology, BAU Mymensingh, India. 2006.
- [18] Muselin F, Trif A, Brezovan D, Stancu A. and Snejana P. The consequences of chronic exposure to lead on liver, spleen, lungs and kidney arhitectonics in rats. *Lucrări Stiinlifice Med. Vet.*, 2010 (2):123-127.
- [19] El-nager D and Aldahmash B. Effect of corn oil, flaxseed oil and black seed oil on testiculardamage induced by lead acetate in albino mice: A histological study. *Pakistan J. Zool.*, 2013; 45:1083-1089.
- [20] Jin Xu, Lian L, Chen Wu, Wang X, Wen Fu, Xu J. Lead induces oxidative stress, DNA damage and alteration of p53, Bax and Bcl-2 expressions in mice. *Food and Chemical Toxicology* 2008; 46:5; 1488–94.
- [21] European Commission. Directive 2010/63/EU of the European Parliament and the Council on the Protection of Animals used for Scientific Purposes. Brussels, Belgium: European Commission, 2010.
- [22] OECD. The organization of Economic Co-operation Development Guideline for Testing of Chemical usage in Experimental Animals. Paris, 2001; pp 1-14.
- [23] Alarifi S, Aldahmash B, EL-Nagar D, Dkhil M. Effect of corn oil, flaxseed oil and black seed oil on lead acetate-induced hepatic tissue damage: A histological study. J. med. Pl. Res., 2012; 6: 4128-4134.
- [24] Sasidharan S, Chen Y, Saravanan D, Sundram KM and Yoga LL. Extraction, Isolation and Characterization of Bioactive Compounds from Plants'Extracts. *Afr J Tradit Complement Altern Med.* 2011, 8(1):1-10
- [25] Schoenwolf GC, Bleyl SB, Brauer PR, Francis-west PH and Philipa H. Larsen's Human Embryology (5<sup>th</sup> ed); 2015, Chapt. 16. New York; Endinburgh: Churchill Livingstone.
- [26] Ekanem AU, Kwari HD, Garba SH, Salami HA. Effect of Lead Acetate on Spleen and Blood Parameters in Albino Rats. *IOSR Journal of Dental and Medical Sciences*; 2015 (14)3, 43-49 www.iosrjournals.org DOI: 10.9790/0853-14314349 www.iosrjournals.org.
- [27] Allouche L, Hamadouche M, Touabti A, Khennouf S. Effect of long-term exposure to low or moderate lead concentrations on growth, lipid profile and liver function in albino rats. *Adv. Biol. Res.* 2011; 5, 339–347.
- [28] Kowolenko M, Tracy L, Mudzinski S, Lawrence D. Effect of lead on macrophage function. *J. Leukoc. Biol.*, 1988; **3**:357-364.
- [29] Aldahmash AB and El-Nager DM. The Protective Effect of Vitamin C Against Toxicity Induced by Lead-Acetate on Liver and Spleen in Swiss Albino Mice. *Pakistan J. Zool.*, 2014; 46(5), 1425-1431.
- [30] Mauel J, Ransijn A, Buchmuller Y. Lead inhibits intracellular killing of leishmania parasites and extracellular cytolysis of target cells by macrophages exposed to macrophage activating factor. *J. Leukoc. Biol.*, 1989; **45**: 401-409.
- [31] Tian L. and Lawrence D. Lead inhibits nitric oxide production in vitro by murine splenic macrophages. *Toxicol. appl. Pharmacol.*, 1995; 132:156-163.

[32] Aly F, Donya M, Abo-Zeid M. The protective role of folic acid, vitamin B12 and vitamin C on the mutagenicity of the anticancer drug daunorubicin. *Researcher*, 2009; 1:16-26.