



# THE ROLE OF PHYTOCHEMICALS IN THE MANAGEMENT OF LIFESTYLE DISORDERS

#### Maria M. Adeyemi

Department of Chemistry and Biochemistry, Caleb University, Lagos, Nigeria.

Email: docmaryadeyemi@gmail.com

#### Cite this article:

Maria M. Adeyemi (2024), The Role of Phytochemicals in The Management of Lifestyle Disorders. African Journal of Environment and Natural Science Research 7(3), 83-99. DOI: 10.52589AJENSR-VJJY3S6C

#### **Manuscript History**

Received: 16 May 2024 Accepted: 18 Jun 2024 Published: 6 Aug 2024

Copyright 2024 T hA uho(). T his is an perturbative of the terms of C reative ommonA tribution N of ommerciaN D environment of A 0 I international C C B N QND 4 0 I, which permits anyone to share use reproduce and redium provided the original author and source are credited

**ABSTRACT:** *Phytochemicals are natural chemical constituents* or bioactive compounds found in medicinal plants, aromatic plants, vegetables, fruits, leaves, flowers and roots which work in conjunction with nutrients and dietary fibers to act as defense against diseases and to slow the aging process. The medicinal values of plants lie in the abilities of these phytochemicals to produce definite physiological action on the human body. A variety of phytochemicals such as polyphenols, terpenoids, flavonoids, tannins, saponins, glycosides and steroids present in plants and their products are key factors in the treatment of several disorders including lifestyle disorders. Lifestyle disorders, also known as non-communicable diseases, are conditions associated with the way people live and behave. According to the World Health Organization (WHO) fact sheets, lifestyle disorders like obesity, diabetes, cancer, cardiovascular diseases, respiratory and gastrointestinal diseases account for 59 percent of the 56.5 million deaths annually and 45.9 percent of the global burden of disease (WHO, 2021). This review introduced and identified various phytochemicals reported with beneficial effects or roles and their functional classification or subclasses that have been identified to help prevent diseases associated with lifestyle changes.

**KEYWORDS:** Cardiovascular diseases, Medicinal plants, Lifestyle disorders, Obesity, Phytochemicals.



## **INTRODUCTION**

Plants represent an enormous reservoir of biologically active compounds used in the treatment of various ailments from times immemorial. These plant constituents are divided into two groups: primary and secondary constituents, based on their functions in plant metabolism. The primary constituents are common sugars, amino acids, proteins and chlorophyll, while the secondary constituents, which are referred to as phytochemicals, are alkaloids, terpenoids, flavonoids, tannins, phenolic compounds, saponins, cardiac glycosides and many more (Adeyemi et al., 2023).

Phytochemicals are natural chemical constituents or bioactive compounds found in medicinal plants, aromatic plants, vegetables, fruits, leaves, flowers and roots which work in conjunction with nutrients and dietary fibers to act as a defense against diseases and to slow the aging process (Adeyemi et al., 2023). The medicinal values of plants lie in these phytochemicals which produce definite physiological action on the human body. A variety of phytochemicals such as polyphenols, alkaloids, terpenoids, flavonoids, tannins, saponins, glycosides, steroids and proteins present in plants and their products are key factors in the treatment of several disorders including lifestyle disorders. The emergence of phytochemicals with health benefits offers an excellent opportunity to improve health and disease conditions and has received much attention in recent years from scientific community, consumers and health care providers as they are being identified and characterized with scientific evidence to support their concept of health promotion.

Lifestyle disorders, also known as non-communicable diseases, are conditions associated with the way people live and behave. As opposed to infectious diseases, they are caused by lack of physical activity, unhealthy eating, alcohol, substance use disorders and smoking tobacco, which can lead to heart disease, stroke, obesity, type II diabetes, and lung cancer (Mathur et al., 2019). According to the World Health Organization (WHO) fact sheets, lifestyle disorders such as diabetes, obesity, cardiovascular diseases, cancer, osteoporosis, respiratory diseases and gastrointestinal diseases account for 59 percent of the 56.5 million deaths annually and 45.9 percent of the global burden of disease (WHO, 2021). Lifestyle and diet are major factors thought to influence susceptibility to many diseases. Other factors such as drug abuse, tobacco smoking and alcohol drinking as well as lack of exercise are associated with risk of developing certain diseases especially at later life. Lifestyle disorder arising from changes in lifestyle, such as increased affluence and urbanization, is associated with increased onset of chronic diseases such as obesity and cardiovascular disease (Farhud, 2015).

### Phytochemicals and Lifestyle Disorders

### **Phytochemicals for Diabetes**

Phytochemicals derived from plants have been used ethno-medicinally for the prevention and/or treatment of diabetes due to the extended belief of their therapeutic properties and safety (Bacanli et al., 2019). Diabetes mellitus is a chronic metabolic disease characterized by elevated levels of blood glucose, insufficiency in production and action of insulin. It is reported that approximately 80% of diabetic patients rely on herbal medicine for successive treatment (Kifle, 2021). Diabetes mellitus is associated with increased formation of free radicals and decreased antioxidant potential (Gaikwad et al., 2014) and the seventh leading cause of death worldwide (Bacanli et al., 2019; Thent & Latiff, 2018). Medicinal plants' phytochemicals are



used alone or in combination with antidiabetic drugs (Ezuruike & Prieto, 2014) and these phytochemicals present an exciting opportunity for the development of new types of therapeutics for diabetes mellitus (Gaikwad et al., 2019; Krawczyk et al., 2023). Phytochemicals which can exert antioxidant and free radical scavenging activities are suggested to improve the insulin sensitivity. Studies (Chanwitheesuk et al., 2005; Bacanli et al., 2019) have shown that phytochemicals showed protective effects against oxidative stress mediated diseases including diabetes and can prevent the formation of Advanced Glycation End-Products (AGEs) and other diabetic complications associated with high oxidative stress conditions (Rahimi et al., 2005). Antioxidant phytochemicals such as allicin, cinnamic acids, coumarins, diterpenes, flavonoids, lignans, monoterpenes, phenylpropanoids, tannins and triterpenes can be found in all parts of plants like wood, bark, stems, pods, leaves, fruit, roots, flowers, pollen and seeds in high concentrations (Table 1, Figure 1).

# Table 1: Main phytochemicals, their plants of origin and major health effects (Ban Canli et al., 2019)

Phytochemicals Quercetin	Plants of Origin Onion Apple Berries Many nuts Brassica <i>Piper sarmentosum</i> <i>Ficus racemose</i>	Health Effects Antihyperglycemic effects Antioxidant effects Ameliorative effects on diabetic complications
Kaempferol	Ficus racemose	Antihyperglycemic effects Antioxidant effects
Naringenin	Ficus racemose	Antihyperglycemic effects Antioxidant effects
Baicalein	Ficus racemose	Antihyperglycemic effects Antioxidants effects
Glabridin	Glycyrrhiza spp	Antihyperglycemic effects Antioxidants effects
Magniferi	Mango	Antidiabetic effects Anticancer effects Antiviral effects Antiaging effects Antioxidant effects
Momorcharins Momordin Charantin Goyasaponins	Momordica charantia	Antihyperglycemic effects Antioxidant effects Ameliorative effects on diabetic complication
Ginsenosides	Panax ginseng	Antidiabetic effects
Oleanolic acid	Olea europaea	Antihyperglycemic effects
Limonene	<i>Citrus</i> plants	Antihyperglycemic effects Antioxidant effects Ameliorative effects on diabetic complications
Ursolic acid	Malus pumila	Antihyperglycemic effects

Article DOI: 10.52589/AJENSR-VJJY3S6C DOI URL: https://doi.org/10.52589/AJENSR-VJJY3S6C



	Ocimum basilicum Vaccinium spp Vaccinium macrocarpon Olea europaea Origanum vulgare Rosmarinus officinalis Salvia spp	Antioxidant effects Ameliorative effects on diabetic complications
Cinnamic acid	Thymus spp. Blueberry Kiwi Cherry Plum Apple Pear Chicory Artichoke Coffee <i>Cinnamonum cassia</i>	Antihyperglycemic effects Antioxidant effects Ameliorative effects on diabetic complications
Cinnamaldehyde	Cinnamomum cassia	Antihyperglycemic effects Ameliorative effects on diabetic complications
Curcumin	Curcuma longa	Antihyperglycemic effects Antioxidant effects Ameliorative effects on diabetic complications
Resveratrol	Grapes Cranberries Blueberries	Antihypergleemic effects Antioxidant effects Ameliorative effects on
Naringin	Tomatoes Grapefruits <i>Citrus</i> plants	diabetic complications Antihyperglycemic effects Antioxidant effects Ameliorative effects on diabetic complications
Catechins	Cocoa	Antihyperglycemic effects Antioxidant effects Ameliorative effects on diabetic complications





Figure 1: Structures of Important phytochemicals for diabetes management (Gaikwad et al., 2014)



## Phytochemicals for Obesity

Obesity termed as the "New World Syndrome" (Nammi et al., 2004) is considered a global problem by the World Health Organization (WHO, 2014) that over 1.4 billion adults aged twenty and older were overweight, among whom almost 300 million women and more than 200 million men were obese (Seyedan et al., 2015). Also, it is estimated that 58% of the world population will become obese by 2030 (Tremmel et al., 2017). Obesity is increased energy intake than energy expenditure resulting in fat deposition and weight gain to an extent where health may be impaired (Muller et al., 2021; WHO, 2014). Many medications have been employed in the treatment and management of obesity over the years. However, despite the unescapable progression of these disease and the promising results of some drugs on lowering body weight and adjustment of numerous cardio-metabolic factors in the past few years, most of the approved anti-obesity drugs have been withdrawn from the market due to serious side effects (Seyedan et al., 2015; Kang & Park, 2012).

Anti-obesity drugs may operate through catecholamine releasing agents such as amphetamine, phentermine, and related substituted amphetamines (e.g., bupropion) which act as appetite suppressants, increasing body metabolism, interfering with the ability of the body to absorb specific nutrients in food and inhibiting enzymes involved in fat absorption. Development of nutrient digestion and absorption of enzyme inhibitors are considered important strategies in the effort to decrease energy intake via gastrointestinal mechanism. Due to these adverse effects of anti-obesity drugs, it is crucial to discover novel inhibitors derived from natural sources (Birari & Bhutani, 2007; Mohammed et al., 2014), particularly plants containing phytochemicals that are not associated with side effects (Sharma & Kanwar, 2018). The potential sites targeted by these phytochemicals include the brain to alter neural signals related to hunger, the gastrointestinal tract involved in nutrient absorption and adipose tissue that plays a vital role in fat storage and degradation. Dietary phytochemicals have been reported to target different stages of the adipocytes (fat cells) life cycle (Williams et al., 2013). These dietary phytochemicals such as polyphenols acts by inducing apoptosis of fat cells, lipolysis and inhibition of inflammation. Based on the possible mode of action to combat obesity, phytochemicals are classified into six major types: (1) lipase inhibitors, (2) appetite suppressants, (3) energy expenditure regulators (thermogenesis), (4) lipid metabolism regulators, (5) adipocyte differentiation regulators and (6) other phytogenic compounds (Balaji et al., 2015) (Figure 2).





Figure 2: Phytochemicals and the major possible target to contain obesity (Balaji et al., 2015)

Phytochemicals for body weight control strategies could be outlined as: i) inhibition of food intake, by inhibiting orexigenic signals or enhancing anorexigenic signals, and limiting the bioavailability of macronutrients, ii) decrease in the caloric content of foods, by substituting sugar or fat less caloric or less digestible substances, iii) stimulation of energy expenditure (thermogenesis), iv) regulation of nutrient partition between tissues and not favoring the efficiency of fat deposition processes. (Manjula & Suneetha, 2011).

### Phytochemicals with Anti-obesity Potential

Phytochemicals that have been found in plants and appeared to possess anti-obesity properties as depicted in Figure 2 are:

# Flavonoids

Flavonoids are a group of plant metabolites containing 15 carbon (C) atoms. Flavonoids are gaining interest because of their potential role in the prevention of diseases such as obesity, cancer, gastrointestinal, cardiovascular and neurodegenerative diseases (Fernandes et al., 2017). Flavonoids such as quercetin show anti-lipase activity by preventing adipogenesis and inducing cell death in mouse pre-adipocytes (Fang et al., 2008; Zheng et al., 2010); Kaempferol extracted from *Bauhinia sp* leaves reduces hyperglycemia (Jorge et al., 2004); and Isorhamnetin found in *Ginkgo biloba, Hippophae sp, Oenanthe javanica* and *Opuntia ficus-indica* possess anti-obesity activities (Rodriguez-Rodriguez et al., 2015).

# Alkaloids

Alkaloids are a class of naturally occurring secondary metabolites with more than 2000 known compounds, mostly containing basic nitrogen atoms. Alkaloids such as synephrine from *citrus aurantium*; piperine from *Piper nigrum*; piperlongumine from *Piper longum*; and liensinine,



isoliensinine, neferine and nuciferine from *Nelumbo nucifera* have shown potential effects on obesity (Acharya & Shrivastava, 2008). Caffeine and chlorogenic acid are the principal constituents of green coffee bean extract which causes reduction in body mass and body fat due to decrease in the absorption of glucose. Decrease in glucose absorption eventually causes an increase in the utilization of fats reserves, due to the reduced availability of glucose as an energy source (Tajik et al., 2017).

# Phenols

Phenols such as p-coumaric, cafferic, ferulic, cinnamic, ellagic and p-hydroxybenzoic acids have been shown to modulate physiological and molecular pathways that are involved in energy metabolism, adiposity and obesity (Son et al., 2010; Krawczyk et al., 2023). Simple phenolic acids are non-flavonoid phenolic compounds which conjugate with other phytochemicals such as flavonoids, alcohols, hydroxy fatty acids, sterols, and glucosides. Ferulic acid has hypocholesteremic and hypoglycemic effects, and thus, it could be effective in lowering the risk of high fat diet-induced obesity (de Melo et al., 2017; Jin Son et al., 2010). Chlorogenic acid and coumaric acid cause significant inhibition of cell growth and enhance apoptosis.

# **Phytosterols**

Phytosterols are structurally similar to mammalian cell-derived cholesterol (Gupta et al., 2011). Phytosterols exist in both esterified and free alcohol forms. Phytosterols that appear to reduce obesity include diosgenin, campesterol, brassicasterol, sitosterol, stigmasterol and guggulsterone. High intakes of these compounds decrease low density lipoprotein-cholesterol levels. In the intestinal lumen, phytosterols compete with cholesterol for micelle formation and thus effectively inhibit cholesterol absorption (Izar et al., 2011). Their influence on intestinal genes and transcription factors make phytosterols the key regulators in metabolism and cholesterol transport in the expression of liver genes (Jesch et al., 2008).

# Terpenoids

Terpenoids (isoprenoids) are chemically modified terpenes and comprise more than 40,000 compounds of both primary and secondary metabolites. Terpenoids such as gymnemic acid from *Gymnerma sylvestre*, oleanolic acid from *Panax ginseng* and corosolic acid from *Lagestroemia speciose* have potential therapeutic effects on obesity (Osman et al., 2010). Peroxisome Proliferator-Activated Receptors (PPAR) activation attenuates obesity and type-2 diabetes. Geranylgeraniol, farnesol and geraniol terpenoids are ligands with the potential to activate PPAR and dietary lipid sensors that control energy homeostasis, as well as lipid and carbohydrate disorders (Goto et al., 2010).

# Phytochemicals as Pancreatic Lipase Inhibitors

One of the strategies used to combat obesity is by interfering at the gastrointestinal level through the inhibition of specific enzymes like lipase and amylase. Lipase is a digestive enzyme, a subclass of the esterases (triacylglycerol hydrolase E.C. 3.1.1.3) that catalyzes the hydrolysis of ester bonds in water-insoluble lipid substrates. Lipase performs essential roles in digestion, and processing of dietary lipids (e.g., triacylglycerols, fats and oils) to monoglycerides and free fatty acids in humans. The decreased digestion and absorption of ingested fats leads to overall decreased caloric absorption leading to decreased obesity (Tsujita



et al., 2006; Balaji et al., 2015). Currently, there are few drugs which can interact with lipases and inhibit their action. Orlistat's lipase inhibitory activity occurs through a covalent bond at the lipase's active site (serine) and has been associated with serious side effects. Therefore, focus on screening novel side effects-free lipase inhibitors derived from plants and other natural sources are essential. Plant phytochemicals provide pancreatic lipase inhibitors with potential for development into clinical products (Table 2).

#### Table 2: Medicinal plants and their active components with lipase inhibitory activity

Plant Salix masudama Aesculus turbinata Coffea canephora	Used parts Leaf Seed	Active components Polyphenol fraction Aesin/escin Caffeine, chlorogenic acid, neochlorogenic acid, feruloylqurinic acid	References Han et al., 2003 Kimura et al., 2006 Shimoda et al., 2006
Xvlonia aethionica	Fruit	Aqueous extract	Etoundi et al., 2010
Scorodophloeus zenkeri	Husk, Seed	Aqueous extract	Etoundi et al., 2010
Baccharis trimera	Stem	Methanolic extract	Souza et al., 2011
Murrava koenigii (L.)	Spreng leaves	Mahanimbine	Rahul et al., 2010
Aronia melanocarpa (L.)	Water extract	Anthocyanidin	Worsztynowicz et al., 2014
Black berry	Stem bark	Ellagic acid	Lei et al., 2007
Pomegranate leaf		Platicodin saponins	Xu et al., 2005
Platycodi radix	Bark	Ethanolic extract	Kim & Kang 2005
Juniperus communis	Wood	Water extract	C
Illicium religiosum	Rhizomes	Chikusetsu saponins	Han et al., 2005
Panax japonicas	Bark, Seed	Ethanolic extract	Sharma et al., 2005; Moreno et al., 2006
Vitis vinifera	Leaves	Ethanolic extract	Sharma et al., 2005
Cudrania tricuspidata	Seeds	Methanolic extract	Jo et al., 2017
Eisenia bicyclis	Brown algae	Phloroglucinol derivatives	Eom et al., 2013
Opuntia ficus-indica	Fructus	Aqueous extract	Padilla-Camberos et al., 2015
N. nucifera petal	Petals	Methanol extract	Ono et al., 2006
Aframomum melegueta, Spilanthes acmella	-	Crude ethanolic extract	Ekanem et al., 2007
Camellia sinensis, Theaceae	Tea	EGCG	Groove et al., 2012
Salacia reticulate	Nut	(-).4-O- methylepigallocatechi	Kishino et al., 2006
Millottia ninnata	Doult		Harri at al 2012
πιμειμα ρίππατα	Dark	Aqueous extract	паті et al., 2013

African Journal of Environment and Natural Science Research ISSN: 2689-9434 Volume 7, Issue 3, 2024 (pp. 83-99)



Terminatia paniculata	Bark	Ethanolic extract	Mopuri & Meriga, 2014
Oolong tea	Leaves		Zhu et al., 2015
Phragmanthera	Leaves	Methanolic extracts	Adeyemi, 2023; 2024
incana			

#### Phytochemicals and Cardiovascular Disease

Cardiovascular disease accounts for roughly 20% of all deaths per year worldwide in both developed and developing countries (Vasanthi et al., 2012). Lifestyle factors including a diet high in saturated fat, energy and in cholesterol, have a crucial part in the onset of Cardiovascular Disease (CVD) risk. Epidemiological studies examining CVD risks in different populations have observed a positive correlation between elevated levels of low density lipoprotein-cholesterol and development of CVD as well as low levels of high density lipoprotein-cholesterol and CVD (Liu et al., 2023; Hsu et al., 2019; Wen et al., 2019; Saito et al., 2020; Al-Shoabi et al., 2023; Razavi et al., 2024; Ren & Wang, 2023). Consuming a diet rich in natural antioxidants has been associated with prevention from and/or treatment of CVD. Bioactive components of food, which are of special interest, include Vitamins E and C, polyphenols, carotenoids (mainly lycopene and  $\beta$ -carotene), and coenzyme Q10, featured by their antioxidant properties. (Islam et al., 2021). A number of bioactive compounds generally obtained from plants, such as icariin, isoflavones, diosgenin, resveratrol, quercetin, catechin, sulforaphane, tocotrienols and carotenoids, are proven to reduce the risk of cardiovascular diseases and aid cardioprotection (Zheng et al., 2022; Guan et al., 2021). The cardioprotective effects of the various phytochemicals are perhaps due to their antioxidative, antihypercholesterolemic, antiangiogenic, anti-ischemic, inhibition of platelet aggregation and anti-inflammatory activities that reduce the risk of cardiovascular disorders (Al-Shoabi et al., 2023; Vasanthi et al., 2012; Islam et al., 2021; Zeng et al., 2022).

#### **Phytochemicals and Cancer**

Cancer is a condition where the body cells multiply in an uncontrollable manner, one of the leading causes of death worldwide, killing approximately 9.6 million people annually despite significant advancements in its treatment over the past decades (Khatoon et al., 2022). Dietary factors are thought to account for about 30% of cancers as diet is second only to tobacco as a potentially preventable cause of cancer (Zam & Hassan, 2019). The contribution of diet to risk of cancer in developing countries is lower around 20% (Padala et al., 2020). Dietary antimutagens which may provide a means of slowing progression towards cancer have been identified such as certain types of dietary fibers, certain probiotics or dietary phytochemicals acting as antioxidants, which include curcumin, ascorbic acid, vitamin E, various polyphenols and carotenoids (George et al., 2021; Xiao & Bai, 2019). Phytochemicals such as curcumin, resveratrol, tocotrienol, and quercetin have emerged as potential chemosensitizing agents in cancer cells due to their less toxic and multitargeted properties. Preclinical and clinical studies enumerated their potential to prevent drug resistance and sensitize cancer cells to chemotherapeutic agents by modulating several genes/proteins or pathways that regulate the key factors during the growth and progression of tumors such as inhibition of anti-apoptotic proteins, activation of pro-apoptotic proteins, reduced expression of different transcription factors, chemokines, enzymes, cell adhesion molecules, protein tyrosine kinases, and cell cycle regulators (George et al., 2021; Khatoon et al., 2022).





Role of Phytochemicals in Cancer Chemoprevention: Insights

Figure 3: Role of phytochemicals in cancer chemoprevention: Insights (George et al., 2021)

### REFERENCES

- Acharya, D. & Shrivastava, A. (2008). Indigenous herbal medicines: Tribal formulations and traditional herbal practices. Aavishar Publishers Distributors, Jaipur.
- Adeyemi, M. M, Shokunbi, O. S. & Osilesi, O. (2023). Qualitative And Quantitative Phytochemical Analysis Of Methanol Extracts Of Phragmanthera Incana (Schum) Leaves Parasitized On Southwest-Nigeria Host Trees. *Current Trends in Life Science Research*, 1: 125–133.
- Adeyemi, M. M. (2023). GC-MS ANALYSIS OF METHANOL EXTRACTS OF Phragmanthera incana LEAVES FROM GUAVA, MANGO, CASHEW AND KOLANUT TREES. *Medical and Health Sciences European Journal*, 7(1). Retrieved from https://aspjournals.org/Journals/index.php/mhsej/article/view/236
- Adeyemi, M. M. (2024). Anti-Obesity potentials of methanol extracts of Phragmanthera incana leaves Hemi-parasitic on guava, cashew, Kolanut and mango trees in high-fat diet-induced obese rats. *African Journal of Biology and Medical Research*, 7(1), 85-94.
- Al-Shoaibi, A. A., Li, Y., Song, Z., Chiang, C., Hirakawa, Y., Saif-Ur-Rahman, K. M., Shimoda, M., Nakano, Y., Matsunaga, M., Aoyama, A., Tamakoshi, K., Ota, A., & Yatsuya, H. (2023). Association of Low-Density Lipoprotein Cholesterol with Risk of Coronary Heart Disease and Stroke among Middle-Aged Japanese Workers: An Analysis using Inverse Probability Weighting. *Journal of atherosclerosis and thrombosis*, 30(5), 455–466. https://doi.org/10.5551/jat.63519
- Bacanli, M., Dilsiz, S. A., Başaran, N., & Başaran, A. A. (2019). Effects of phytochemicals against diabetes. *Advances in food and nutrition research*, *89*, 209–238. https://doi.org/10.1016/bs.afnr.2019.02.006
- Balaji, M., Ganjayi, M. S., Hanuma Kumar, G. E., Parim, B. N., Mopuri, R., & Dasari, S. (2016). A review on possible therapeutic targets to contain obesity: The role of



phytochemicals. *Obesity research & clinical practice*, *10*(4), 363–380. https://doi.org/10.1016/j.orcp.2015.12.004

- Birari, R. B., & Bhutani, K. K. (2007). Pancreatic lipase inhibitors from natural sources: unexplored potential. *Drug discovery today*, *12*(19-20), 879–889. https://doi.org/10.1016/j.drudis.2007.07.024
- Birari, R., Javia, V., & Bhutani, K. K. (2010). Antiobesity and lipid lowering effects of Murraya koenigii (L.) Spreng leaves extracts and mahanimbine on high fat diet induced obese rats. *Fitoterapia*, 81(8), 1129–1133. https://doi.org/10.1016/j.fitote.2010.07.013
- Chanwitheesuk, A., Teerawutgulrag, A., & Rakariyatham, N. (2005). Screening of antioxidant activity and antioxidant compounds of some edible plants of Thailand. *Food Chemistry*, *92*, 491-497.
- de Melo, T. S., Lima, P. R., Carvalho, K. M., Fontenele, T. M., Solon, F. R., Tomé, A. R., de Lemos, T. L., da Cruz Fonseca, S. G., Santos, F. A., Rao, V. S., & de Queiroz, M. G. (2017). Ferulic acid lowers body weight and visceral fat accumulation via modulation of enzymatic, hormonal and inflammatory changes in a mouse model of high-fat diet-induced obesity. *Brazilian journal of medical and biological research = Revista brasileira de pesquisas medicas e biologicas*, 50(1), e5630. https://doi.org/10.1590/1414-431X20165630
- Ekanem, A. P., Wang, M., Simon, J. E., & Moreno, D. A. (2007). Antiobesity properties of two African plants (Afromomum meleguetta and Spilanthes acmella) by pancreatic lipase inhibition. *Phytotherapy* research : *PTR*, 21(12), 1253–1255. https://doi.org/10.1002/ptr.2239
- Eom, S. H., Lee, M. S., Lee, E. W., Kim, Y. M., & Kim, T. H. (2013). Pancreatic lipase inhibitory activity of phlorotannins isolated from Eisenia bicyclis. *Phytotherapy research* : *PTR*, 27(1), 148–151. https://doi.org/10.1002/ptr.4694
- Etoundi CB, Kuate D, Ngondi JL, Oben J. Anti-amylase, anti-lipase and antioxidant effects of aqueous extracts of some Cameroonian spices. Journal of Natural Product 2010; 3:165–71.
- Ezuruike, U. F., & Prieto, J. M. (2014). The use of plants in the traditional management of diabetes in Nigeria: pharmacological and toxicological considerations. *Journal of ethnopharmacology*, 155(2), 857–924. https://doi.org/10.1016/j.jep.2014.05.055
- Fang, X. K., Gao, J., & Zhu, D. N. (2008). Kaempferol and quercetin isolated from Euonymus alatus improve glucose uptake of 3T3-L1 cells without adipogenesis activity. *Life sciences*, 82(11-12), 615–622. https://doi.org/10.1016/j.lfs.2007.12.021
- Farhud D. D. (2015). Impact of Lifestyle on Health. *Iranian journal of public health*, 44(11), 1442–1444.
- Fernandes, I., Pérez-Gregorio, R., Soares, S., Mateus, N., & de Freitas, V. (2017). Wine Flavonoids in Health and Disease Prevention. *Molecules (Basel, Switzerland)*, 22(2), 292. https://doi.org/10.3390/molecules22020292
- Gaikwad, S. B, Mohan, G. K, Rani, M. S. (2014). Phytochemicals for diabetes management. *Pharmaceutical Crops*, 5(sup 1: M2): 11–28.
- George, B. P., Chandran, R. & Abrahamse, H. (2021). Role of phytochemicals in cancer chemoprevention: Insights. *Antioxidants*, 14;10(9):1455.
- Goto, T., Takahashi, N., Hirai, S., & Kawada, T. (2010). Various Terpenoids Derived from Herbal and Dietary Plants Function as PPAR Modulators and Regulate Carbohydrate and Lipid Metabolism. *PPAR research*, 2010, 483958. https://doi.org/10.1155/2010/483958
- Grove, K. A., Sae-tan, S., Kennett, M. J., & Lambert, J. D. (2012). (-)-Epigallocatechin-3gallate inhibits pancreatic lipase and reduces body weight gain in high fat-fed obese

African Journal of Environment and Natural Science Research

ISSN: 2689-9434

Volume 7, Issue 3, 2024 (pp. 83-99)



mice. Obesity (Silver Spring, Md.), 20(11), 2311–2313. https://doi.org/10.1038/oby.2011.139

- Guan, R., Van Le, Q., Yang, H., Zhang, D., Gu, H., Yang, Y., Sonne, C., Lam, S. S., Zhong, J., Jianguang, Z., Liu, R., & Peng, W. (2021). A review of dietary phytochemicals and their relation to oxidative stress and human diseases. *Chemosphere*, 271, 129499. https://doi.org/10.1016/j.chemosphere.2020.129499
- Gupta, A. K., Savopoulos, C. G., Ahuja, J., & Hatzitolios, A. I. (2011). Role of phytosterols in lipid-lowering: current perspectives. QJM : monthly journal of the Association of Physicians, 104(4), 301–308. https://doi.org/10.1093/qjmed/hcr007
- Han, L. K., Sumiyoshi, M., Zheng, Y. N., Okuda, H., & Kimura, Y. (2003). Anti-obesity action of Salix matsudana leaves (Part 2). Isolation of anti-obesity effectors from polyphenol fractions of Salix matsudana. *Phytotherapy research : PTR*, 17(10), 1195–1198. https://doi.org/10.1002/ptr.1405
- Han, L. K., Zheng, Y. N., Yoshikawa, M., Okuda, H., & Kimura, Y. (2005). Anti-obesity effects of chikusetsusaponins isolated from Panax japonicus rhizomes. *BMC complementary and alternative medicine*, 5, 9. https://doi.org/10.1186/1472-6882-5-9
- Hari Venkatesh, K.R. & Chethana, G. S. (2013). Antilipase activity: comparative study of Operculina turpethum, Milletia pinnata (bark and seed) with reference to Curcuma longa. *Journal of Obesity, 1*(1):0–22.
- Hsu, S. H. J., Jang, M. H., Torng, P. L., & Su, T. C. (2019). Positive association between small dense low-density lipoprotein cholesterol concentration and biomarkers of inflammation, thrombosis, and prediabetes in non-diabetic adults. *Journal of Atherosclerosis and Thrombosis*, 26(7), 624–635.
- Islam, S. U., Ahmed, M. B., Ahsan, H., & Lee, Y. S. (2021). Recent Molecular Mechanisms and Beneficial Effects of Phytochemicals and Plant-Based Whole Foods in Reducing LDL-C and Preventing Cardiovascular Disease. *Antioxidants (Basel, Switzerland)*, 10(5), 784. https://doi.org/10.3390/antiox10050784
- Izar, M. C., Tegani, D. M., Kasmas, S. H., & Fonseca, F. A. (2011). Phytosterols and phytosterolemia: gene-diet interactions. *Genes & nutrition*, 6(1), 17–26. https://doi.org/10.1007/s12263-010-0182-x
- Jesch, E. D., Lee J. Y., & Carr, T. P. (2008). Dietary plant sterols regulate genes involved in cholesterol metabolism in mouse liver but not intestine. *Federation of American Societies of Experimental Biology Journal*, 22: 700–735.
- Jin Son, M., W Rico, C., Hyun Nam, S., & Young Kang, M. (2010). Influence of oryzanol and ferulic Acid on the lipid metabolism and antioxidative status in high fat-fed mice. *Journal* of clinical biochemistry and nutrition, 46(2), 150–156. https://doi.org/10.3164/jcbn.09-98
- Jo, Y. H., Kim, S. B., Liu, Q., Do, S. G., Hwang, B. Y., & Lee, M. K. (2017). Comparison of pancreatic lipase inhibitory isoflavonoids from unripe and ripe fruits of Cudrania tricuspidata. *PloS one*, 12(3), e0172069. https://doi.org/10.1371/journal.pone.0172069
- Jorge, A. P., Horst, H., de Sousa, E., Pizzolatti, M. G., & Silva, F. R. (2004). Insulinomimetic effects of kaempferitrin on glycaemia and on 14C-glucose uptake in rat soleus muscle. *Chemico-biological interactions*, *149*(2-3), 89–96. https://doi.org/10.1016/j.cbi.2004.07.001
- Kang, J. G., & Park, C. Y. (2012). Anti-Obesity Drugs: A Review about Their Effects and Safety. *Diabetes & metabolism journal*, 36(1), 13–25. https://doi.org/10.4093/dmj.2012.36.1.13



- Khatoon, E., Banik, K., Harsha, C., Sailo, B. L., Thakur, K. K., Khwairakpam, A. D., Vikkurthi, R., Devi, T. B., Gupta, S. C., & Kunnumakkara, A. B. (2022). Phytochemicals in cancer cell chemosensitization: Current knowledge and future perspectives. *Seminars in cancer biology*, 80, 306–339. https://doi.org/10.1016/j.semcancer.2020.06.014
- Kifle, Z. D., Bayleyegn, B., Yimer Tadesse, T., & Woldeyohanins, A. E. (2021). Prevalence and associated factors of herbal medicine use among adult diabetes mellitus patients at government hospital, Ethiopia: An institutional-based cross-sectional study. *Metabolism* open, 11, 100120. https://doi.org/10.1016/j.metop.2021.100120
- Kim, H.Y & Kang, M. H. (2005). Screening of Korean medicinal plants for lipase inhibitory activity. Phytotherapy Research, 19: 359–61.
- Kimura, H., Ogawa, S., Jisaka, M., Kimura, Y., Katsube, T., & Yokota, K. (2006). Identification of novel saponins from edible seeds of Japanese horse chestnut (Aesculus turbinata Blume) after treatment with wooden ashes and their nutraceutical activity. *Journal of pharmaceutical and biomedical analysis*, 41(5), 1657–1665. https://doi.org/10.1016/j.jpba.2006.02.031
- Kishino, E., Ito, T., Fujita, K., & Kiuchi, Y. (2006). A mixture of the Salacia reticulata (Kotala himbutu) aqueous extract and cyclodextrin reduces the accumulation of visceral fat mass in mice and rats with high-fat diet-induced obesity. *The Journal of nutrition*, *136*(2), 433–439. https://doi.org/10.1093/jn/136.2.433
- Krawczyk, M., Burzynska-Pedziwiatr, I., Wozniak, L. A., & Bukowiecka-Matusiak, M. (2023). Impact of Polyphenols on Inflammatory and Oxidative Stress Factors in Diabetes Mellitus: Nutritional Antioxidants and Their Application in Improving Antidiabetic Therapy. *Biomolecules*, 13(9), 1402. https://doi.org/10.3390/biom13091402
- Lei, F., Zhang, X. N., Wang, W., Xing, D. M., Xie, W. D., Su, H., & Du, L. J. (2007). Evidence of anti-obesity effects of the pomegranate leaf extract in high-fat diet induced obese mice. *International journal of obesity (2005)*, 31(6), 1023–1029. https://doi.org/10.1038/sj.ijo.0803502
- Liu, H., Li, J., Liu, F., Huang, K., Cao, J., Chen, S., Li, H., Shen, C., Hu, D., Huang, J., Lu, X., & Gu, D. (2023). Efficacy and safety of low levels of low-density lipoprotein cholesterol: trans-ancestry linear and non-linear Mendelian randomization analyses. *European journal of preventive cardiology*, 30(12), 1207–1215. https://doi.org/10.1093/eurjpc/zwad111
- Manjula, K & Suneetha, C. (2011). Designer Foods- Their role in preventing Lifestyle Disorders. *International Journal of Natural Science*, 2(4): 878-882.
- Mathur, P., & Mascarenhas, L. (2019). Life style Diseases: *Keeping Fit for a Better Tomorrow. The Indian journal of medical research*, *149*(Suppl), S129–S135. https://doi.org/10.4103/0971-5916.251669
- Mohammed, G. A, Ibrahim, S. R, El Kahayat, E., El Dine, R. (2014). Natural Anti-Obesity agents. *Bulletin of Faculty of Pharmacy, Cairo University, 52*(2): 269–284.
- Mopuri, R, & Meriga, B. (2014). Anti-lipase and anti-obesity activities of Terminalia paniculata bark in high calorie dietinduced obese rats. *Global Journal of Pharmacology 8*(1):114-9, <u>http://dx.doi.org/10.5829/idosi.gjp.2014.8.1.82221</u>.
- Moreno, D. A, Ilic, N, Poulev, A., & Raskin, I. (2006). Effects of Arachis hypogaea nutshell extract on lipid metabolic enzymes and obesity parameters. *Life Sciences*, 78:2797-803.
- Müller, T. D., Blüher, M., Tschöp, M. H., & DiMarchi, R. D. (2022). Anti-obesity drug discovery: advances and challenges. *Nature reviews. Drug discovery*, 21(3), 201–223. https://doi.org/10.1038/s41573-021-00337-8



- Ono, Y., Hattori, E., Fukaya, Y., Imai, S., & Ohizumi, Y. (2006). Anti-obesity effect of Nelumbo nucifera leaves extract in mice and rats. *Journal of ethnopharmacology*, 106(2), 238– 244. https://doi.org/10.1016/j.jep.2005.12.036
- Osman, M, Fayed, S. A, Ghada, I. M, Romeliah, R. M. (2014). Protective effects of chitosan ascorbic acid and Gymnema sylvestre against hypercholesterolemia in male rats. *Australian Journal of Basic Applied Sciences*, 20104: 89–98.
- Padala SA, Barsouk A, Thandra KC, Saginala K, Mohammed A, Vakiti A, Rawla P, Barsouk A. Epidemiology of renal cell carcinoma. World journal of oncology. 2020 Jun;11(3):79.
- Padilla-Camberos, E., Flores-Fernandez, J. M., Fernandez-Flores, O., Gutierrez-Mercado, Y., Carmona-de la Luz, J., Sandoval-Salas, F., Mendez-Carreto, C., & Allen, K. (2015). Hypocholesterolemic Effect and In Vitro Pancreatic Lipase Inhibitory Activity of an Opuntia ficus-indica Extract. *BioMed research international*, 2015, 837452. https://doi.org/10.1155/2015/837452
- Rahimi, R., Nikfar, S., Larijani, B., & Abdollahi, M. (2005). A review on the role of antioxidants in the management of diabetes and its complications. *Biomedicine & pharmacotherapy* = *Biomedecine & pharmacotherapie*, 59(7), 365–373. https://doi.org/10.1016/j.biopha.2005.07.002
- Razavi, A. C., Jain, V., Grandhi, G. R., Patel, P., Karagiannis, A., Patel, N., Dhindsa, D. S., Liu, C., Desai, S. R., Almuwaqqat, Z., Sun, Y. V., Vaccarino, V., Quyyumi, A. A., Sperling, L. S., & Mehta, A. (2024). Does Elevated High-Density Lipoprotein Cholesterol Protect Against Cardiovascular Disease?. *The Journal of clinical endocrinology and metabolism*, 109(2), 321–332. https://doi.org/10.1210/clinem/dgad406
- Ren, X., & Wang, X. (2023). Association of the low-density lipoprotein cholesterol to highdensity lipoprotein cholesterol ratio and major adverse cardiac and cerebrovascular events in patients with coronary heart disease undergoing percutaneous coronary intervention: a cohort study. *Current medical research and opinion*, 39(9), 1175–1181. https://doi.org/10.1080/03007995.2023.2246889
- Rodriguez-Rodriguez, C., Torres, N., Guiterrez-Uribe, J. A, Noriega, L. G, Torre-Villalvazo, I. (2015). The effect of isorhamnetin glycosides extracted from Opuntia ficus-indica in a mouse model of diet induced obesity. *Food Functionality*, 6: 805–815.
- Saito, I., Yamagishi, K., Kokubo, Y., Yatsuya, H., Iso, H., Sawada, N., Inoue, M., & Tsugane, S. (2020). Non-High-Density Lipoprotein Cholesterol and Risk of Stroke Subtypes and Coronary Heart Disease: The Japan Public Health Center-Based Prospective (JPHC) Study. *Journal of atherosclerosis and thrombosis*, 27(4), 363–374. https://doi.org/10.5551/jat.50385
- Seyedan, A., Alshawsh, M. A., Alshagga, M. A., Koosha, S., & Mohamed, Z. (2015). Medicinal Plants and Their Inhibitory Activities against Pancreatic Lipase: A Review. *Evidencebased complementary and alternative medicine : eCAM*, 2015, 973143. https://doi.org/10.1155/2015/973143
- Sharma, N., Sharma, V. K., & Seo, S. Y. (2005). Screening of some medicinal plants for antilipase activity. *Journal of ethnopharmacology*, 97(3), 453–456. https://doi.org/10.1016/j.jep.2004.11.009
- Sharma, T, & Karmar, S. S. (2018). Phytomolecules for Obesity and Body weight management. *Journal of Biochemistry and cell Biology*, 1:1.
- Shimoda, H., Seki, E., & Aitani, M. (2006). Inhibitory effect of green coffee bean extract on fat accumulation and body weight gain in mice. *BMC complementary and alternative medicine*, *6*, 9. https://doi.org/10.1186/1472-6882-6-9



- Souza, S. P. d., Pereira, L. L. S., Souza, A. A., & Santos, C. D. d. (2011). Inhibition of pancreatic lipase by extracts of baccharis trimera (less.) dc., asteraceae: evaluation of antinutrients and effect on glycosidases. Revista Brasileira De Farmacognosia, 21(3), 450-455. https://doi.org/10.1590/s0102-695x2011005000049
- Tajik, N., Tajik, M., Mack, I., & Enck, P. (2017). The potential effects of chlorogenic acid, the main phenolic components in coffee, on health: a comprehensive review of the literature. *European journal of nutrition*, 56(7), 2215–2244. https://doi.org/10.1007/s00394-017-1379-1
- Thent, Z. C. & Latiff, A. A. (2018). Savior of Diabetes: Antioxidants in diabetes food plan. IntechOpen.
- Tremmel, M., Gerdtham, U. G., Nilsson, P. M., & Saha, S. (2017). Economic Burden of Obesity: A Systematic Literature Review. *International journal of environmental research and public health*, 14(4), 435. https://doi.org/10.3390/ijerph14040435
- Tsujita, T., Takaichi, H., Takaku, T., Aoyama, S., & Hiraki, J. (2006). Antiobesity action of epsilon-polylysine, a potent inhibitor of pancreatic lipase. *Journal of lipid* research, 47(8), 1852–1858. https://doi.org/10.1194/jlr.M600168-JLR200
- Vasanthi, H. R., ShriShriMal, N., & Das, D. K. (2012). Retraction Notice: Phytochemicals from plants to combat cardiovascular disease. *Current medicinal chemistry*, 19(14), 2242– 2251. https://doi.org/10.2174/092986712800229078 (Retraction published Curr Med Chem. 2020;27(32):5444)
- Wen, J., Huang, Y., Lu, Y., & Yuan, H. (2019). Associations of non-high-density lipoprotein cholesterol, triglycerides and the total cholesterol/HDL-c ratio with arterial stiffness independent of low-density lipoprotein cholesterol in a Chinese population. *Hypertension* research, 42(8), 1223–1230.
- WHO (2008). Traditional medicines. http://www.who.int/mediacentre/factsheets//134/en.
- WHO 2021. Cardiovascular diseases. https://www.who.int/news-room/factsheets/detail/cardiovascular-diseases-(cvds)
- Williams DJ, Edwards D, Hamernig I, Jian L, James AP, Johnson SK, Tapsell LC (2013) Vegetables containing phytochemicals with potential anti-obesity properties. A Rev. Food Res Int. 2013;52: 323–333.
- World Health Organization, (2014). "Obesity and overweight,",<u>http://www.who.int/mediacentre/factsheets/fs311/en/</u>.
- Worsztynowicz P, Napierała M, Białas W, Grajek W, Olkowicz M. (2014). Pancreatic -amylase and lipase inhibitory activity of polyphenolic compounds present in the extract of black chokeberry (Aronia melanocarpa L.). Process Biochem 49(9):1457–63.
- Xiao, J & Bai, W. (2019). Bioactive phytochemicals. *Critical Reviews in Food Science and Nutrition*, 26;59(6):827–9.
- Xu, B. J, Han, L. K, Zheng, Y. N, Lee, J. H, Sung, C. K. (2005). In vitro inhibitory effect of triterpenoidal saponins from Platycodi radix on pancreatic lipase. Archive of Pharmaceutical Research 28:180–5.
- Zam, W. & Hassan, B. (2021) Diet influence on colorectal cancer. *Progress in nutrition, 1*;21(2-S):42–8.
- Zeng, Y, Xiong, Y, Yang, T, Wang, Y, Zeng, J, Zhou, S, Luo, Y, & Li, L. (2022). Icariin and its metabolites as potential protective phytochemicals against cardiovascular disease: From effects to molecular mechanisms. *Biomedicine & Pharmacotherapy* 1;147:112642.
- Zheng, C. D., Duan, Y. Q., Gao, J. M., & Ruan, Z. G. (2010). Screening for anti-lipase properties of 37 traditional Chinese medicinal herbs. *Journal of the Chinese Medical Association : JCMA*, 73(6), 319–324. https://doi.org/10.1016/S1726-4901(10)70068-X





Zhu, Y. T., Ren, X. Y., Yuan, L., Liu, Y. M., Liang, J., & Liao, X. (2015). Fast identification of lipase inhibitors in oolong tea by using lipase functionalised Fe3O4 magnetic nanoparticles coupled with UPLC-MS/MS. *Food chemistry*, 173, 521–526. https://doi.org/10.1016/j.foodchem.2014.10.087